

BONE-GRAFT SUBSTITUTES: *FACTS, FICTIONS &* *APPLICATIONS*

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A REALITY CHECK

It is estimated that more than 500,000 bone-grafting procedures are performed annually in the United States, with approximately half of these procedures related to spine fusion. These numbers easily double on a global basis and indicate a shortage in the availability of musculoskeletal donor tissue traditionally used in these reconstructions. (Figure 1)

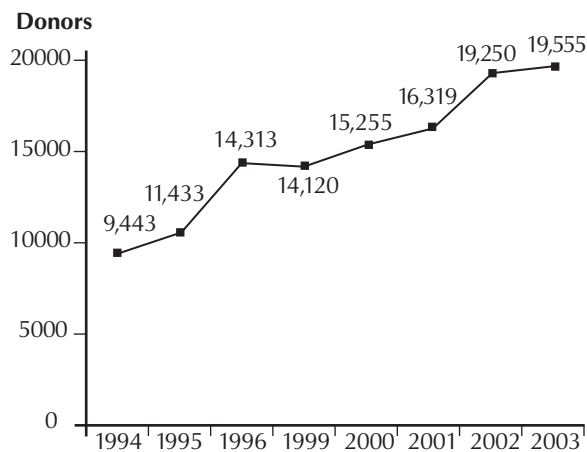


Figure 1: U.S. trends in musculoskeletal tissue donors. Source: AATB Annual Survey

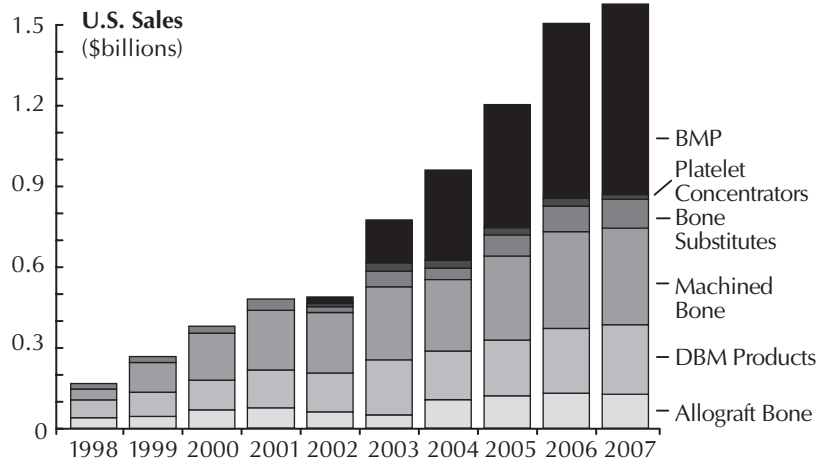


Figure 2: U.S. sales of bone graft and bone-graft substitutes Source: Orthopedic Network News

This reality has stimulated a proliferation of corporate interest in supplying what is seen as a growing market in bone replacement materials. (Figure 2) These graft alternatives are subjected to varying degrees of regulatory scrutiny, and thus their true effectiveness in patients may not be known prior to their use by orthopaedic surgeons. It is important to gain insight into this emerging class of bone-graft alternatives.

THE PHYSIOLOGY OF BONE GRAFTING

The biology of bone grafts and their substitutes is appreciated from an understanding of the bone formation processes of Osteogenesis, Osteoinduction and Osteoconduction.

Graft Osteogenesis: The cellular elements within a donor graft, which survive transplantation and synthesize new bone at the recipient site.

Graft Osteoinduction: New bone realized through the active recruitment of host mesenchymal stem cells from the surrounding tissue, which differentiate into bone-forming osteoblasts. This process is facilitated by the presence of growth factors within the graft, principally bone morphogenetic proteins (BMPs).

Graft Osteoconduction: The facilitation of a bone healing process into a defined passive trellis structure.

All bone graft and bone-graft substitute materials can be described through these processes.

While fresh autologous graft has the capability of supporting new bone growth by all three means, it may not be necessary for a bone graft replacement to inherently have all three properties in order to be clinically effective. When inductive molecules are locally delivered on a scaffold, mesenchymal stem cells are ultimately attracted to the site and are capable of reproducibly inducing new bone formation, provided minimal concentration and dose thresholds are met. In some clinical studies, osteoinductive agents have been shown to potentially perform equivalent or superiorly to autograft demonstrating efficacy as an autograft replacement.

However, bone marrow aspirate applied to osteoconductive scaffolds are still reliant on the local mechanical and biological signals in order to ultimately form bone. For this reason, these materials are typically used as an adjunct in order to retain efficacy equivalent to autograft.

Similarly, osteoconductive materials work well when filling non-critical size defects that would normally heal easily. However, in more challenging critical size defects, either fresh autologous bone graft or osteoinductive agents appear necessary for healing.

BONE AUTOGRAFTS

Fresh autogenous cancellous and, to a lesser degree, cortical bone are benchmark graft materials that allograft and bone substitutes attempt to match in *in vivo* performance. They incorporate all of the mentioned properties, are harvested at both primary and secondary surgical sites, and have no associated risk of viral transmission. Furthermore, they offer structural support to implanted devices and, ultimately, become mechanically efficient structures as they are incorporated into surrounding bone through creeping substitution. However, the availability of autografts is limited and harvest is often associated with donor-site morbidity.

BONE ALLOGRAFTS

The advantages of bone allograft recovered from deceased donor sources include its ready availability in various shapes and sizes, avoidance of the need to sacrifice host structures, and there is no donor-site morbidity. Bone allografts are distributed through regional tissue banks and by most major orthopaedic and spinal companies. Still, the grafts are not without controversy, particularly regarding their association with the transmission of infectious agents. Some tissue processors incorporate methods that may eliminate the risk. However, uncontrolled and unvalidated processing and irradiation protocols may alter graft biomechanical and biochemical properties. It is critical to know your tissue bank provider to ensure their processing and preservation methods inactivate viruses but do not negatively alter the biomechanical and biochemical properties of the tissues intended for a particular clinical use. A comparison of properties of allograft and autograft bone is shown in Figure 3. Often, in complex surgical reconstructions, these materials are used in tandem with implants and fixation devices. (Figure 4)

| Bone Graft | Structural Strength | Osteo-Conduction | Osteo-Induction | Osteogenesis |
|---|---------------------|------------------|-----------------|--------------|
| Autograft | | | | |
| Cancellous | No | +++ | +++ | +++ |
| Cortical | +++ | ++ | ++ | ++ |
| Allograft | | | | |
| <i>Cancellous</i> | | | | |
| Frozen | No | ++ | + | No |
| Freeze-Dry | No | ++ | + | No |
| <i>Cortical</i> | | | | |
| Frozen | +++ | + | No | No |
| Freeze-Dry | + | + | No | No |
| Demineralized Allogeneic Cancellous Chips | No | + | ++ | No |

Figure 3: Comparative properties of bone grafts

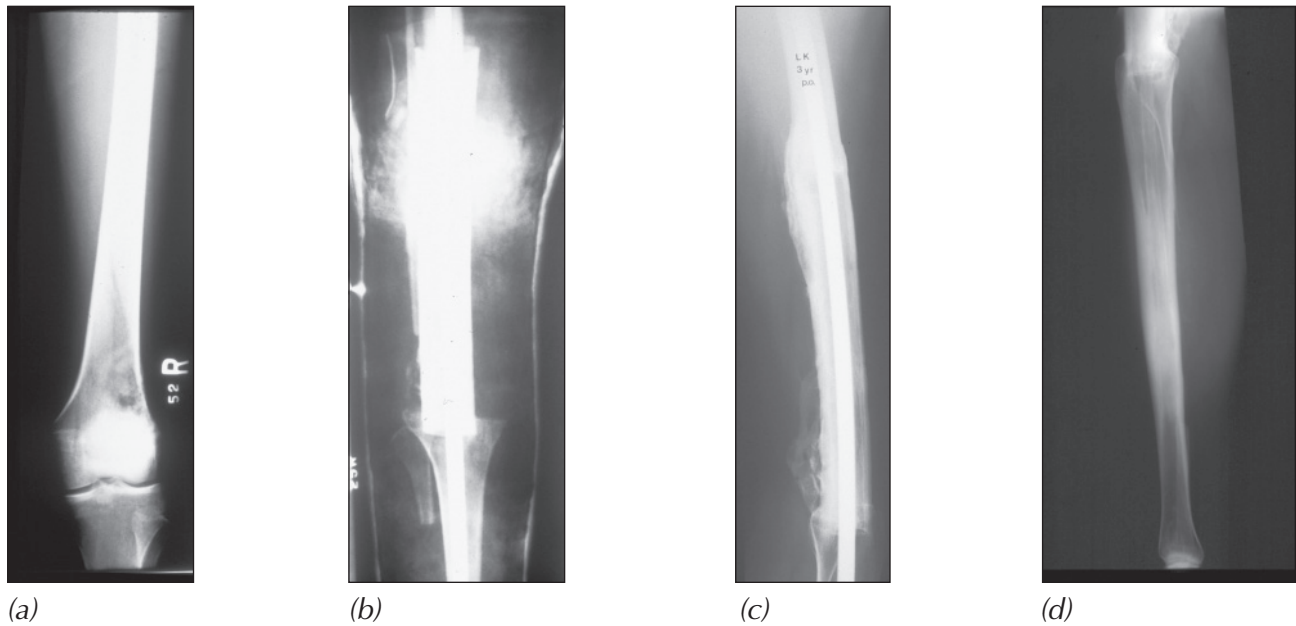


Figure 4: (a) A 17-year old patient with osteosarcoma of the distal femur with no extraosseous extension or metastatic disease. Following chemotherapy, (b) limb salvage with wide resection was performed. Femoral reconstruction with the use of an autogenous cortical fibular graft, iliac crest bone chips, morselized cancellous autograft and structural allograft combined with internal fixation. (c) Graft incorporation and remodeling are seen at 3 years. (d) Limb restoration is noted at 10 years following resection. (The intramedullary rod was removed at 5 years.)

BONE GRAFT SUBSTITUTES

The ideal bone-graft substitute is biocompatible, bioresorbable, osteoconductive, osteoinductive, structurally similar to bone, easy to use, and cost-effective. Within these parameters a growing number of bone graft alternatives are commercially available for orthopaedic applications, including reconstruction of cavitory bone deficiency and augmentation in situations of segmental bone loss and spine fusion. They are variable in their composition and their claimed mechanisms of action. A series of case examples demonstrate their mechanisms of action through the healing process. (Figures 5-8)

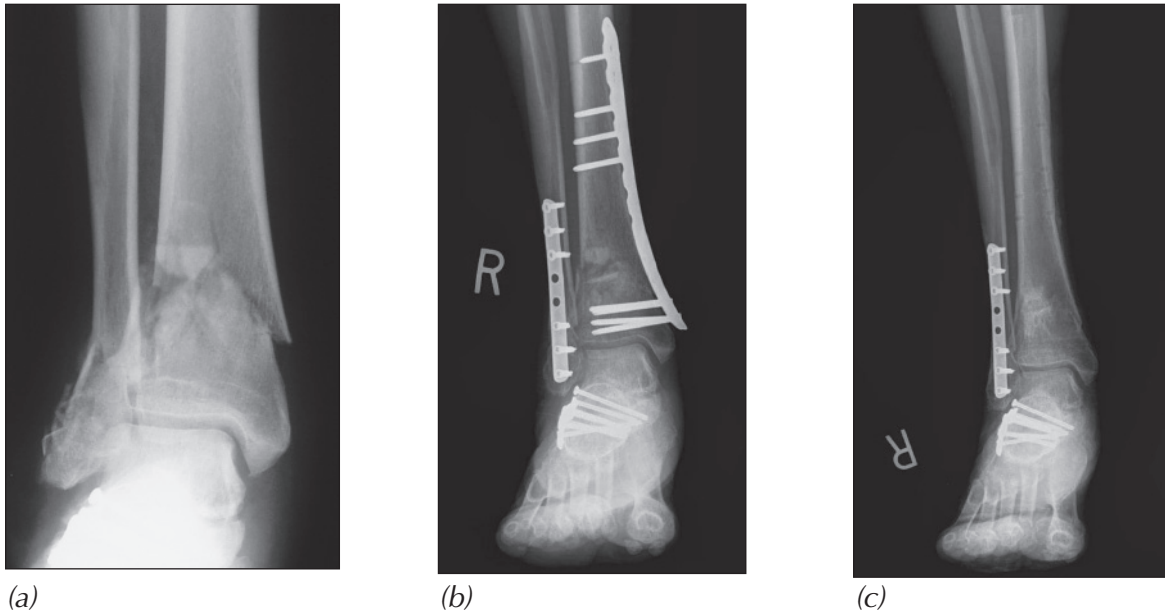


Figure 5: (a) A 61-year old male with a comminuted pilon fracture sustained in a motor vehicle accident. (b) After 2 months with an external fixator, definitive fixation of the tibia with a percutaneous injection of IGNITE® (Wright Medical Technology, Inc., Arlington, TN) graft to bridge the slow-healing fracture. (c) Two years post-op, the fracture is consolidated and the patient is ambulating pain-free.

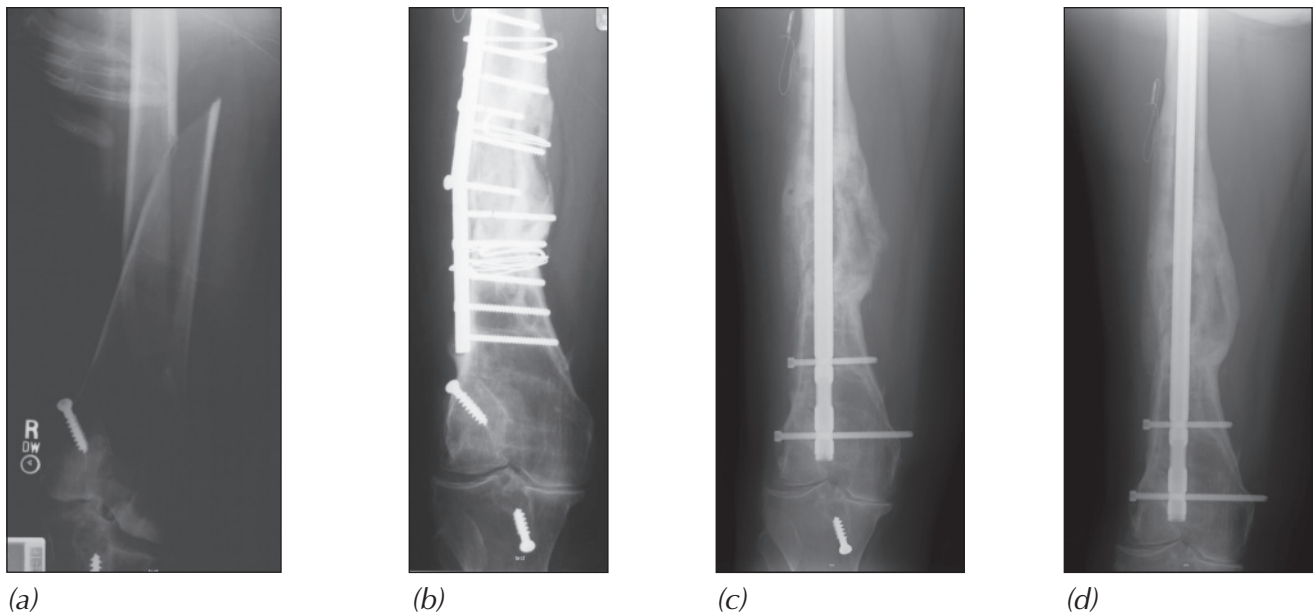


Figure 6: (a) A 58-year old obese female with a nonunion of the right femur after falling from a horse. Treatments included plating with cortical struts and DBM. (b) Nine months following third surgery the plate and several screws are broken. (c) Three months after treatment with IM rod fixation and OP-1® Implant (Stryker Biotech, Hopkinton, MA) she was full weight bearing, with full range of motion and pain free. (d) Nine months postoperative.

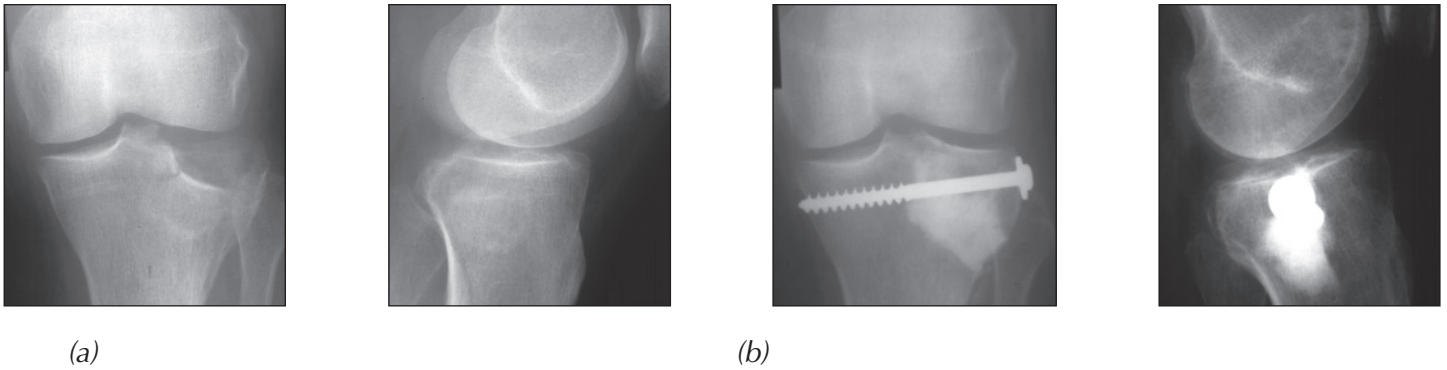


Figure 7: (a) AP and Lateral radiographs, 67-year old female with depressed fracture of the lateral tibial plateau. (b) AP and Lateral radiographs 12 months after ORIF with filling the defect with Norian® SRS® (Synthes USA, Paoli, PA). No loss of reduction of the plateau surface is noted, fracture completely healed.

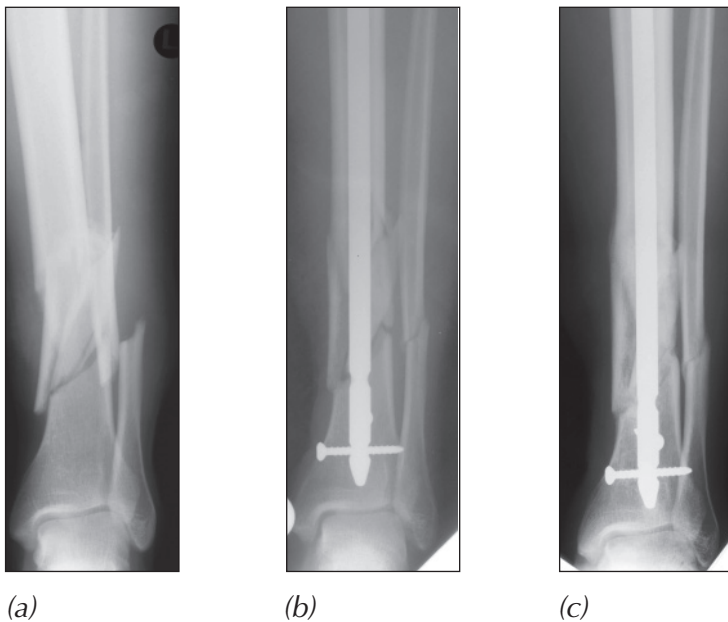


Figure 8: (a) A 29-year old male with a Grade III B oblique fracture of the distal tibia from a motorcycle accident. (b) Six weeks after being treated with an unreamed locked nail and INFUSE® Bone Graft (Medtronic Spinal & Biologics, Memphis, TN). (c) Patient full weight bearing and radiographically healed 20 weeks post-operative.

BURDEN OF PROOF

It is reasonable to assume that not all bone-graft alternatives will perform the same. This presents a challenging choice for the orthopaedic surgeon. As a first principle, it is important to appreciate that different healing environments (e.g., a metaphyseal defect, a long-bone fracture, an interbody spine fusion, or a posterolateral spine fusion) have different levels of difficulty in forming new bone. For example, a metaphyseal defect will permit the successful use of many purely osteoconductive materials. In contrast, a posterolateral spine fusion will not succeed if purely osteoconductive materials are used as a stand-alone substitute. Thus, validation of any bone-graft alternative in one clinical site may not necessarily predict its performance in another location.

A second principle is to seek the highest burden of proof reported from clinical and preclinical studies to justify the use of an osteoinductive graft material or the choice of one alternative over another. It is generally more difficult to make bone in humans than it is in larger order animals. Only human trials can determine the efficacy of bone-graft substitutes in humans as well as their site-specific effectiveness. In this latter context, surgeons should practice evidence-based medicine and tailor treatment for patients based on the published medical literature and the levels of evidence claimed. (Wright JG, Swiontkowski MF, Heckman JD. *Introducing levels of evidence to the journal. J Bone Joint Surg Am.* 2003 Jan;85(1):1-3.)

BURDEN OF PROOF (Cont'd.)

A third principle requiring burden of proof specifically pertains to products that are not subjected to high levels of regulatory scrutiny, such as 100% demineralized bone matrix (DBM) or platelet gels containing “autologous growth factors.” Such products are considered to involve minimal manipulation of cells or tissue and are thus regulated as tissue rather than as devices. When DBM products include additives, they require 510(k) clearance. As a result, there is no standardized level of proof of safety and effectiveness required before these products are marketed and are used in patients. While these products may satisfy regulatory requirements, testing in relevant animal models is limited or absent and there is a risk that they will not produce the expected results in humans.

FUTURE

FDA approvals include the use of PMA approved rhBMP-2 (INFUSE® Bone Graft) as an autograft replacement in spinal fusion and treatment of open tibia fractures; rhBMP-7 (OP-1® Implant) is Humanitarian Use Device (HDE) approved as an autograft substitute for long bone nonunions; and rhBMP-7 (OP-1® Putty) is HDE approved as an alternative to autograft in compromised patients requiring revision posterolateral (intertransverse) lumbar spinal fusion, for whom autologous bone and bone marrow recovery are not feasible or are not expected to promote fusion. These clinical applications demonstrate impressive osteoinductive capacity and pave the way for broader clinical applications. Their methods of administration include direct placement in the surgical site, but results have been more promising when the growth factors have been administered in combination with substrates to facilitate timed-release delivery and/or provide a material scaffold for bone formation. FDA regulatory imperatives will continue to determine their availability. Their cost/benefit ratio will ultimately influence clinical use.

Further advances in tissue engineering, “the integration of the biological, physical and engineering sciences,” will create new carrier constructs that regenerate and restore tissue to its functional state. These constructs are likely to encompass additional families of growth factors, evolving biological scaffolds and incorporation of mesenchymal stem cells. Ultimately, the development of *ex vivo* bioreactors capable of bone manufacture with the appropriate biomechanical cues will provide tissue-engineered constructs for direct use in the skeletal system.

TAKE HOME MESSAGE

- The increasing number of bone-grafting procedures performed annually in the U.S. has created a shortage of cadaver allograft material and a need to increase musculoskeletal tissue donation.
 - This has stimulated corporate interest in developing and supplying a rapidly expanding number of bone-graft substitutes, the makeup of which includes natural, synthetic, human and animal-derived materials.
 - Fresh autogenous cancellous and, to a lesser degree, cortical bone are the benchmark graft materials. Their shortcomings include limited availability and donor-site morbidity.
 - The advantages of allograft bone include availability in various sizes and shapes as well as avoidance of host structure sacrifice and donor-site morbidity. Transmission of infection, particularly the human immunodeficiency virus (HIV) has been virtually eliminated as a concern. The properties of the allograft should be confirmed with the tissue provider to ensure they correspond with their intended clinical use.
 - The ideal bone-graft substitute is biocompatible, bioresorbable, osteoconductive, osteoinductive, structurally similar to bone, easy to use, and cost-effective. Currently marketed products are variable in their composition and their claimed mechanisms of action. It is reasonable that not all bone-graft substitute products will perform the same.
 - FDA approvals for specific uses of recombinant human growth factors (rhBMP-2 (INFUSE® Bone Graft) and rhBMP-7 (OP-1® Implant and OP-1® Putty)) are based on demonstrated bone repair in human trials. Other applications will likely emerge.
 - The orthopaedic surgeon has many choices for bone grafting. *Caveat emptor!* Selection should be based on reasoned burdens of proof. These include examination of the product claims and whether they are supported by preclinical and human studies in site-specific locations where they are to be utilized in surgery. It is imperative to appreciate the level of evidence claimed in the latter studies.
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Summary of typical bone-graft substitutes that are commercially available - 2008

| Company | Commercially available product | Composition | Commercially available forms | Claimed mechanisms of action | Burdens of proof | FDA status |
|-----------------------|--------------------------------|--|---|--|---|--|
| AlloSource | AlloFuse™ | Heat sensitive copolymer with DBM | Injectable gel and putty | <ul style="list-style-type: none"> Osteoconduction Bioresorbable Osteoinduction | <ul style="list-style-type: none"> Case reports Animal studies Cell culture | <ul style="list-style-type: none"> 510(k) cleared |
| Biomet Osteobiologics | ProOsteon® 500R | Coralline-derived hydroxyapatite/calcium carbonate (HA/CC) composite | Granular or block | <ul style="list-style-type: none"> Osteoconduction Bioresorbable | <ul style="list-style-type: none"> Human studies Case reports Animal studies | <ul style="list-style-type: none"> 510(k) cleared |
| | InterGro® | DBM in a lecithin carrier | Paste, putty and mix with HA/CC composite granules | <ul style="list-style-type: none"> Osteoconduction Bioresorbable Osteoinduction | <ul style="list-style-type: none"> Case reports Animal studies Every lot tested for osteoinduction | <ul style="list-style-type: none"> 510(k) cleared |
| | BonePlast® | Calcium sulfate with or without HA/CC composite granules | Various volumes of powder and setting solution | <ul style="list-style-type: none"> Osteoconduction Bioresorbable | <ul style="list-style-type: none"> Case reports Animal studies | <ul style="list-style-type: none"> 510(k) cleared |
| DePuy Spine | HEALOS® Bone Graft Replacement | Mineralized collagen matrix | Variety of strip sizes | <ul style="list-style-type: none"> Osteoconduction Creeping substitution Osteoinduction Osteogenesis when mixed with bone marrow aspirate | <ul style="list-style-type: none"> Peer-reviewed, published human studies Case reports Animal studies | <ul style="list-style-type: none"> 510(k) cleared |
| | CONDUIT® TCP Granules | 100% β-TCP | Granules | <ul style="list-style-type: none"> Osteoconduction Bioresorbable | <ul style="list-style-type: none"> Case reports Animal studies | <ul style="list-style-type: none"> 510(k) cleared |
| Exactech | Opteform® | DBM and cortical cancellous chips suspended in gelatin carrier | Formable putty or dry powder ready to be hydrated with autologous diluents or saline | <ul style="list-style-type: none"> Osteoconduction Bioresorbable Osteoinduction Osteogenesis when mixed with autogenous bone graft | <ul style="list-style-type: none"> Human studies Case reports Animal studies Every lot tested <i>in vivo</i> for osteoinduction | <ul style="list-style-type: none"> 510(k) cleared |
| | Optefil® | DBM suspended in gelatin carrier | Injectable bone paste, dry powder ready to be hydrated with blood or saline | <ul style="list-style-type: none"> Osteoconduction Bioresorbable Osteoinduction Osteogenesis when mixed with autogenous bone graft | <ul style="list-style-type: none"> Human studies Case reports Animal studies Every lot tested <i>in vivo</i> for osteoinduction | <ul style="list-style-type: none"> 510(k) cleared |
| | Optecure® | DBM suspended in a hydrogel carrier | Dry mix kit delivered with or buffered saline mix with patient's autogenous bone graft or autologous diluents | <ul style="list-style-type: none"> Osteoconduction Bioresorbable Osteoinduction Osteogenesis when mixed with autogenous bone graft | <ul style="list-style-type: none"> Human studies Case reports Animal studies Every lot tested <i>in vivo</i> for osteoinduction | <ul style="list-style-type: none"> 510(k) cleared |
| | Optecure® + CCC | DBM and CCC suspended in a hydrogel carrier | Dry mix kit delivered with buffered saline or mix with patient's autogenous bone graft or autologous diluents | <ul style="list-style-type: none"> Osteoconduction Bioresorbable Osteoinduction Osteogenesis when mixed with autogenous bone graft | <ul style="list-style-type: none"> Animal studies Every lot tested <i>in vivo</i> for osteoinduction | <ul style="list-style-type: none"> 510(k) cleared |
| | OpteMx™ | HA/TCP biphasic combination | Granules, sticks, rounded wedges, wedges and cylinders in several sizes | <ul style="list-style-type: none"> Osteoconduction Bioresorbable Compressive strength of 400psi Osteogenesis and limited osteoinduction when mixed with bone marrow aspirate | <ul style="list-style-type: none"> Human studies Case reports Animal studies | <ul style="list-style-type: none"> 510(k) cleared |

| Company | Commercially available product | Composition | Commercially available forms | Claimed mechanisms of action | Burdens of proof | FDA status |
|--|--------------------------------|--|---|---|--|--|
| Integra Orthobiologics/ (IsoTis Orthobiologics) | Accell 100™ | DBM putty | Injectable putty | <ul style="list-style-type: none"> Osteoconduction Bioresorbable Osteoinduction | <ul style="list-style-type: none"> Human studies Case reports Animal studies Every DBM lot tested for osteoinduction | <ul style="list-style-type: none"> 510(k) cleared |
| | Accell Connexus® | DBM plus reverse phase medium | Injectable putty | <ul style="list-style-type: none"> Osteoconduction Bioresorbable Osteoinduction | <ul style="list-style-type: none"> Human studies Case reports Animal studies Every DBM lot tested for osteoinduction | <ul style="list-style-type: none"> 510(k) cleared |
| | Accell TBM™ | Total bone matrix, 100% preformed | Various sized strips | <ul style="list-style-type: none"> Osteoconduction Bioresorbable Osteoinduction | <ul style="list-style-type: none"> Human studies Case reports Animal studies Every DBM lot tested for osteoinduction | <ul style="list-style-type: none"> 510(k) cleared |
| | Integra Mozaik™ | <ul style="list-style-type: none"> 80% highly purified B-TCP 20% highly purified type-1 collagen | Strip and putty | <ul style="list-style-type: none"> Osteoconduction Bioresorbable | <ul style="list-style-type: none"> Animal studies Case reports | <ul style="list-style-type: none"> 510(k) cleared |
| LifeNet Health | Optium DBM® | DBM combined with glycerol carrier | Formable putty (bone fibers) and injectable gel (bone particles) | <ul style="list-style-type: none"> Osteoconduction Bioresorbable Osteoinduction | <ul style="list-style-type: none"> Human studies Case reports Animal studies | <ul style="list-style-type: none"> 510(k) cleared |
| | IC Graft Chamber® | DBM particles and cancellous chips | Lyophilized and packaged in various sizes within a delivery chamber | <ul style="list-style-type: none"> Osteoconduction Bioresorbable Osteoinduction Designed to be used with blood, PRP or bone marrow to enhance DBM activity. | <ul style="list-style-type: none"> Animal studies Case reports | <ul style="list-style-type: none"> Regulated under CFR 1270 and 1271 as a human tissue and 510(k) cleared |
| | Cellect DBM® | DBM fibers and cancellous chips | Provided in a specialized cartridge | <ul style="list-style-type: none"> Osteoconduction Bioresorbable Osteoinduction Designed for the retention of osteoprogenitor cells | <ul style="list-style-type: none"> Animal studies Case reports | <ul style="list-style-type: none"> Regulated under CFR 1270 and 1271 as a human tissue and 510(k) cleared |
| Medtronic Spinal & Biologics | INFUSE® Bone Graft | rhBMP-2 protein on an absorbable collagen sponge | Multiple kit sizes | <ul style="list-style-type: none"> Bioresorbable carrier Osteoinduction Chemotaxis of stem cells; indirect osteogenesis | <ul style="list-style-type: none"> Human studies (Level I and Level III data) Case reports Animal studies | <ul style="list-style-type: none"> PMA approved for fusion with spinal cage PMA approved for open tibia fractures with IM nail |
| | MasterGraft® Granules | Biphasic calcium phosphate | Granules | <ul style="list-style-type: none"> Osteoconduction Bioresorbable | <ul style="list-style-type: none"> Animal studies | <ul style="list-style-type: none"> 510(k) cleared |
| | MasterGraft® Matrix | Calcium phosphate and collagen | Compression resistant block | <ul style="list-style-type: none"> Osteoconduction Bioresorbable | <ul style="list-style-type: none"> Animal studies | <ul style="list-style-type: none"> 510(k) cleared |
| | MasterGraft® Putty | Calcium phosphate and collagen | Moldable putty | <ul style="list-style-type: none"> Osteoconduction Bioresorbable | <ul style="list-style-type: none"> Animal studies | <ul style="list-style-type: none"> 510(k) cleared |

| Company | Commercially available product | Composition | Commercially available forms | Claimed mechanisms of action | Burdens of proof | FDA status |
|------------------------------|--------------------------------|---|--|--|---|---|
| Medtronic Spinal & Biologics | Progenix™ DBM Putty | DBM in Type-1 bovine collagen and sodium alginate | Ready to use injectable putty | <ul style="list-style-type: none"> Osteoconduction Bioresorbable Osteoinduction | <ul style="list-style-type: none"> Animal studies | <ul style="list-style-type: none"> 510(k) cleared |
| | Osteofil® DBM | DBM in porcine gelatin | Injectable paste and moldable strips | <ul style="list-style-type: none"> Osteoconduction Bioresorbable Osteoinduction | <ul style="list-style-type: none"> Animal studies Case reports | <ul style="list-style-type: none"> 510(k) cleared |
| MTF/Synthes | DBX® | DBM in sodium hyaluronate carrier | Paste, putty mix and strip | <ul style="list-style-type: none"> Osteoconduction Bioresorbable Osteoinduction | <ul style="list-style-type: none"> Human studies Case reports Animal studies | <ul style="list-style-type: none"> 510(k) cleared |
| NovaBone/MTF | NovaBone® | Bioactive silicate | Particulate and putty | <ul style="list-style-type: none"> Osteoconduction Bioresorbable Osteostimulation | <ul style="list-style-type: none"> Case reports Animal studies | <ul style="list-style-type: none"> 510(k) cleared |
| Orthovita | Vitoss® | 100% β-TCP and 80% β-TCP/20% collagen | Putty, strip, flow, morsels and shapes | <ul style="list-style-type: none"> Osteoconduction Bioresorbable | <ul style="list-style-type: none"> Published human studies Case reports Animal studies | <ul style="list-style-type: none"> 510(k) cleared |
| Osteotech | Grafton® | DBM combined with Glycerol | Formable putty, injectable gel, putty mixed with chips, flexible sheets and matrix | <ul style="list-style-type: none"> Osteoconduction Bioresorbable Osteoinduction | <ul style="list-style-type: none"> Published human studies Case reports Animal studies | <ul style="list-style-type: none"> 510(k) cleared |
| | Grafton Plus® | DBM combined with a starch carrier | Paste | <ul style="list-style-type: none"> Osteoconduction Bioresorbable Osteoinduction | <ul style="list-style-type: none"> Case reports Animal studies | <ul style="list-style-type: none"> 510(k) cleared |
| Regeneration Technologies | BioSet™ | DBM combined with natural gelatin carrier | Injectable paste, injectable putty, strips and blocks with cortical cancellous chips | <ul style="list-style-type: none"> Osteoconduction Bioresorbable Osteoinduction | <ul style="list-style-type: none"> Human studies Case reports Animal studies Every lot tested <i>in vivo</i> for osteoinduction | <ul style="list-style-type: none"> 510(k) cleared |
| Smith & Nephew | VIAGRAF | DBM combined with glycerol | Putty, paste, gel, crunch and flex | <ul style="list-style-type: none"> Osteoconduction Bioresorbable Osteoinduction | <ul style="list-style-type: none"> Animal studies | <ul style="list-style-type: none"> 510(k) cleared |
| Stryker Biotech | OP-1® Implant | rhBMP-7 with type 1 bone collagen | Lyophilized powder reconstituted to form wet sand | <ul style="list-style-type: none"> Bioresorbable scaffold Osteoinduction | <ul style="list-style-type: none"> Human studies (Level I data) Case reports Animal studies | <ul style="list-style-type: none"> HDE approval for long bone nonunions |
| | OP-1® Putty | rhBMP-7 with type 1 bone collagen | Lyophilized powder reconstituted to form putty | <ul style="list-style-type: none"> Bioresorbable scaffold Osteoinduction | <ul style="list-style-type: none"> Human studies (Level I data) Case reports Animal studies | <ul style="list-style-type: none"> HDE approval for revision posterolateral fusion |
| | Calstrux™ | Tricalcium phosphate with carboxy-methylcellulose | Moldable putty | <ul style="list-style-type: none"> Osteoconduction Bioresorbable | <ul style="list-style-type: none"> Animal studies | <ul style="list-style-type: none"> 510(k) cleared |

| Company | Commercially available product | Composition | Commercially available forms | Claimed mechanisms of action | Burdens of proof | FDA status |
|---------------------------|----------------------------------|--|---|--|---|--|
| Synthes | Norian® SRS® | Calcium phosphate | Injectable paste | <ul style="list-style-type: none"> Osteoconduction Bioresorbable | <ul style="list-style-type: none"> Human studies Case reports Animal studies | • 510(k) cleared |
| | Norian® SRS® Fast Set Putty | Calcium phosphate | Moldable putty | <ul style="list-style-type: none"> Osteoconduction Bioresorbable | <ul style="list-style-type: none"> Human studies Case reports Animal studies | • 510(k) cleared |
| | chronOS® | β-tricalcium phosphate | Granules, blocks and wedges | <ul style="list-style-type: none"> Osteoconduction Bioresorbable | <ul style="list-style-type: none"> Animal studies | • 510(k) cleared |
| | Calceon® 6 | Calcium sulfate | Pellets | <ul style="list-style-type: none"> Osteoconduction Bioresorbable | <ul style="list-style-type: none"> Animal studies | • 510(k) cleared |
| Wright Medical Technology | OSTEOSET® | Surgical grade calcium sulfate | Various sized pellets | <ul style="list-style-type: none"> Osteoconduction Bioresorbable | <ul style="list-style-type: none"> Human studies Case reports Animal studies | • 510(k) cleared |
| | MIIG® X3 | High strength surgical grade calcium sulfate | Minimally invasive injectable graft for compression fractures | <ul style="list-style-type: none"> Osteoconduction Bioresorbable | <ul style="list-style-type: none"> Human studies Case reports Animal studies | • 510(k) cleared |
| | CELLPLEX® | Tricalcium phosphate | Various sized granules | <ul style="list-style-type: none"> Osteoconduction Bioresorbable | <ul style="list-style-type: none"> Case reports Animal studies | • 510(k) cleared |
| | ALLOMATRIX® | DBM with/without CBM in surgical grade calcium sulfate powder | Various volumes of injectable/ formable putty | <ul style="list-style-type: none"> Osteoconduction Bioresorbable Osteoinduction | <ul style="list-style-type: none"> Human studies Case reports Animal studies Cell culture | • 510(k) cleared |
| | ALLOMATRIX® RCS | DBM with CACIPLEX™ Technology in surgical grade calcium sulfate powder | Various volumes of formable putty | <ul style="list-style-type: none"> Osteoconduction Bioresorbable Osteoinduction | <ul style="list-style-type: none"> Animal studies | • 510(k) cleared |
| | IGNITE® | DBM in surgical grade calcium sulfate powder to be mixed with bone marrow aspirate | Percutaneous graft for problem fractures | <ul style="list-style-type: none"> Osteoconduction Bioresorbable Osteoinduction | <ul style="list-style-type: none"> Human studies Case reports Animal studies Cell culture | • 510(k) cleared |
| Zimmer | CopiOs® Bone Void Filler | Dibasic calcium phosphate and Type I collagen | Sponge and paste | <ul style="list-style-type: none"> Osteoconduction Bioresorbable Osteogenesis and osteoinduction when mixed with bone marrow aspirate | <ul style="list-style-type: none"> Case reports Animal studies | • 510(k) cleared |
| | CopiOs® Cancellous Bone Graft | Bovine bone | Cancellous chips, cancellous cubes and wedges | <ul style="list-style-type: none"> Osteoconduction | <ul style="list-style-type: none"> Case reports Animal studies | • 510(k) cleared |
| | Puros® Demineralized Bone Matrix | DBM putty | Putty | <ul style="list-style-type: none"> Osteoconduction Bioresorbable Osteoinduction | <ul style="list-style-type: none"> Every lot tested <i>in vivo</i> for osteoinduction | <ul style="list-style-type: none"> 100% derived from allograft tissue FDA clearance not required |